



## Review Article

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### OTOACOUSTIC EMISSIONS IN TYPE 2 DIABETES: A SYSTEMATIC REVIEW

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#### ABSTRACT

Outer row of hair cells in the organ of Corti are not directly involved in deciding the threshold of the acoustic stimulus, but their damage will increase the hearing threshold and may even cause the deafness. Type 2 diabetes is increasing globally at an alarming rate, which is affecting the normal hearing. In type 2 diabetes poor metabolic control, neuropathy or microangiopathy may affect the normal hearing. This study is focused on self enhancement of the knowledge in the particular field apart from providing the complete review on effect of type 2 diabetes on otoacoustic emissions to the readers. Articles published in English on otoacoustic emissions in type 2 diabetes were collected and reviewed from Pubmed, Google and Google Scholar. Hearing threshold is increased in type 2 diabetic patients and amplitude of otoacoustic emissions was decreased. Hearing loss is observed in type 2 diabetes and it is attributed to damage of outer row of hair cells in organ of Corti.

**Keywords:** otoacoustic emissions, type 2 diabetes, outer row of hair cells

#### INTRODUCTION

Otoacoustic emissions are low level acoustic signals originated from the outer row of hair cells of the cochlea<sup>1,2</sup> and they are used widely to study the cochlear function and hair cell micromechanics in humans<sup>3</sup>. Outer row of hair cells play a significant role in normal hearing when the sound stimulus intensity is below 60 dB<sup>4</sup> and they give an indirect idea about the functional status of inner hair cells<sup>5</sup>. Otoacoustic emissions are useful in screening for cochlear (outer hair cell) functioning, differentiation of cochlear versus retro cochlear auditory dysfunction and also in diagnosis of auditory dysfunction. Cellular metabolism in the outer row of hair cells increases dramatically when they are activated and the outer row of hair cells rapidly elongate during hyper-polarization and become shorter during depolarization. Change in outer hair cell length generates energy within the cochlea that contributes to hearing sensitivity and the ability to distinguish small differences in the frequencies of sounds<sup>6</sup>. Transient evoked otoacoustic emissions are not recorded with audiometric thresholds greater than 25 to 30 dB HL<sup>7</sup>. The absence of distortion product otoacoustic emissions with normal middle ear function is an indication of audiometric thresholds greater than 30 to 35 dB HL<sup>8</sup>. Globally 382 million people have diabetes and the number is set to rise beyond 592 million in less than 25 years. Currently 175 million undiagnosed diabetic cases are present across the globe and the number is increasing at a robust rate, a vast amount of people with diabetes are progressing towards complications unawares<sup>9</sup>. Like all the living cells, neuronal cells that are involved in auditory system also requires glucose for their survival and also for complex signaling process. And this suggests that the cochlea may also be a target organ for the damaging effects caused by hyperglycemia in type 2 diabetes<sup>10</sup>. In type 2 diabetes, hyperglycemia causes widespread tissue damage, most specifically injuring neural tissue<sup>11, 12</sup>. Type 2 diabetes effects

extracellular matrix and results in thickening of the collagen fibers<sup>13 - 15</sup>. In diabetes, protein kinase C is up regulated and binds with PX2 receptors on mammalian hair cells, activating a down regulation of Na/K/ATPase causing elevated extracellular K<sup>+</sup> and intracellular Na<sup>+</sup> and Ca<sup>++</sup> and resulting in excitotoxicity<sup>16, 17</sup>. This study is focused on self enhancement of the knowledge in the particular field for the authors apart from providing the complete review of articles on effect of type2 diabetes on otoacoustic emissions to the readers. Published articles in English on otoacoustic emissions in type 2 diabetes were collected and reviewed from Pubmed, Google and Google Scholar. Exclusion criteria: Otoacoustic emissions in type 1 diabetes were excluded.

#### OBSERVATION & CONCLUSION

Cochlear function is affected to different degrees at different frequency levels in type 2 diabetic patients<sup>18</sup>. Mean amplitudes of otoacoustic emissions were decreased in type 2 diabetes induced CAB/CaJ mouse model for lower and higher frequencies<sup>19</sup>. Hearing loss observed in type2 diabetes induced rhesus monkeys is predominantly cochlea<sup>20</sup>. Decreased amplitude of distortion product otoacoustic emissions were observed in type 2 diabetic patients when compared with the control subjects<sup>21 - 27</sup>. Right ear outer row of hair cells are damaged more than the left ear outer row of hair cells in type 2 diabetic patients<sup>10</sup>. Ears with damaged outer row of hair cells have reduced sensitivity as well as broader tuning than ears with normal outer row of hair cells<sup>28 - 31</sup>. Brain metabolites are also altered in type 2 diabetes; this may also have contributed for the hearing changes in diabetics<sup>32</sup>. All the studies have shown that hyperglycemia in type 2 diabetes decreases the amplitude of outer row of hair cells. They also have shown compelling evidence on increased hearing threshold in type 2 diabetics. Auditory screening with otoacoustic emissions is useful in early

detection of increased hearing threshold and decreased amplitude of outer row of hair cells, which is very essential in taking the prophylactic measures for preventing the further damage. So routine screening for hearing in type 2 diabetic patients with distortion product otoacoustic emissions is advisable. Longitudinal cohort studies are required on otoacoustic emissions for drawing the correlation between the hyperglycemia and functioning of outer row of hair cells in type 2 diabetics.

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#### REFERENECS

- Kemp DT. Stimulated acoustic emissions from within the human auditory system. *J Acoust Soc Am*, 1978; 64: 1386 – 1391.
- Lonsbury-Martin BL, McCoy MJ, Whitehead ML, Martin GK. Clinical testing of distortion-product otoacoustic emissions. *Ear Hear*, 1993; 14: 11–22.
- Berlin, C. Hair Cell Micro-mechanics and Otoacoustic Emissions. Delmar Learning, Thomason Learning, Clifton Park, NY, 2002: 1–155.
- Spoedlin H. The organization of the cochlear receptor. In *Adv Otorhinolaryngol*, 1966; vol 13.
- Gary Rance. Auditory Neuropathy/Dys – synchrony and Its Perceptual Consequences. *Trends in Amplification*, 2005; 9(1): 5.
- James W. Hall III. A Guide to Otoacoustic Emissions (OAEs) for Otolaryngologists. Maico Diagnostics, 2009: 1 – 17.
- Prieve BA et al. Analysis of transient- evoked otoacoustic emissions in normal – hearing and hearing – impaired ears. *J Acoust Soc Am*, 1993; 93:3008 – 3319.
- Gorga MP et al. From laboratory to clinic: a large scale study of distortion product otoacoustic emissions in ears with normal hearing and ears with hearing loss. *Ear Hear*, 1997; 18:440 – 455.
- Leonor Guariguata et al. International Diabetes Federation Diabetes Atlas, 2013; 6<sup>th</sup> Ed: 66. [www.idf.org/diabetesatlas](http://www.idf.org/diabetesatlas). Accessed 12/10/2014.
- Susan T. Frisina et al. Characterization of hearing loss in aged type II diabetes. *Hearing Research*, 2006; 211: 103 – 113.
- Cameron NE, Cotter MA, Low PA. Nerve blood flow in early diabetes in rats: relation to conduction deficits. *Am J Physiol* 1991; 261:e1–e8.
- Stevens MJ, Lattimer SA, Feldman EL, et al. Acetyl-L carnitine deficiency as a cause of altered nerve myo-inositol content, Na<sup>+</sup>/K<sup>+</sup>/ATPase and motor conduction velocity in the streptozocin diabetic rat. *Metabol Clin Exp* 1996; 45:865–872.
- Boyd-White J, Williams JC Jr. Effects of cross-linking on matrix permeability: a model for AGE-modified basement membranes. *Diabetes* 1996; 45:348.
- Charonis AS, Tsilbary EC. Structural and functional changes of laminin and type IV collagen after nonenzymatic glycation. *Diabetes* 1992; 41 (Suppl 2):49–51.
- Haitoglou CS, Tsilbary EC, Brownlee M, Charonis AS. Altered cellular interactions between endothelial cells and non-enzymatically glycosylated laminin /type IV collagen. *J Biol Chem* 1992; 267:12404– 12407.
- Sweeny, G., Klip, A. Regulation of the Na<sup>+</sup>/ K<sup>+</sup> ATPase by insulin: why and how? *Mol. Cell. Biochem*, 1998; 182: 121 – 133.
- Boue – Grabot, E., Archambault, V., Seguela, P. A protein Kinase C site highly conserved in P2X subunits controls the desensitization kinetics of P2XX (2) ATO gated channels. *J. Biol. Chem*, 2000; 275: 10190 – 10195.
- Hayriye Karabulut et al. Evaluation of outer hair cell function and medial olivocochlear efferent system in patients with type II diabetes mellitus. *Turk J Med Sci*, 2014; 44: 150 – 156.
- Vasilyeva ON et al. Interactions of hearing loss and diabetes mellitus in the middle aged CBA/CAJ mouse model of presbycusis. *Hear Res*, 2009 Mar; 249(1 - 2): 44 – 53.
- Fowler CG et al. Hyperinsulinemia / diabetes, hearing and aging in the University of Wisconsin calorie restriction monkeys. *Hear Res*, 2015 Oct; 328:78 – 86.
- Wang H, Zhong N. A study on DPOAE in patients with diabetes mellitus. *Lin Chuang Er Bi Yan Hou Ke Za Zhi*, 1998 Nov; 12(11):4836.
- Park MS, Park SW, Choi JH. Distortion product otoacoustic emissions in diabetics with normal hearing. *Scand Audiol*, 2001; 30(52): 148–151.
- Lisowska G, Namyslowski G, Morawski K, Strojek K. Cochlear dysfunction and diabetic microangiopathy. *Scand Audiol*, 2001; 30(52): 199–203.
- Aladağ I, Kurt S, Eyibilen A, Güven M, Erkorkmaz U. Early evaluation of auditory dysfunction in patients with type 2 diabetes mellitus. *Kulak Burun Bogaz Ihtis Derg*, 2008 Jul-Aug; 18 (4):20310.
- Ren J, Zhao P, Chen L, Xu A, Brown SN, Xiao X. Hearing loss in middle aged subjects with type 2 diabetes mellitus. *Arch Med Res*. 2009 Jan; 40 (1):1823.
- Eren E, Harman E, Arslanoğlu S, Onal K. Effects of Type 2 Diabetes on Otoacoustic Emissions and the Medial Olivocochlear Reflex. *Otolaryngol Head Neck Surg*, 2014 Mar 26; 150 (6):1033-1039.
- Sasso FC et al. Cochlear dysfunction in type 2 diabetes: a complication independent of neuropathy and acute hyperglycemia. *Metabol Clin Exp* 1999; 48:1346–1350.
- Dallos P, Wang CY. Bioelectric correlates of kanamycin intoxication. *Audiology*, 1974; 13:277 – 289.
- Harrison RV, Evans EF. Cochlear fibre responses in guinea pigs with well defined cochlear lesions. *Scand Audiol Suppl*, 1974; 99:30 – 33.
- Ryan AF, Dallos P. Absence of cochlear outer hair cells: effect on behavioural auditory thresholds. *Nature*, 1975; 253:44 – 46.
- Lieberman MC, Dodds LW. Single neuron labelling and chronic cochlear pathology: III. Stereocilia damage and alterations of threshold tuning curves. *Hear Res*, 1984; 16:55 –74.
- Rajani Santhakumari, IndlaYogananda Reddy, Archana R, Rajesh P. Role of yoga in alienating the memory decline and frontal lobe metabolite changes in type 2 diabetes. *Int. J. Res. Ayurveda Pharm.* 2016; 7(1):78-81 <http://dx.doi.org/10.7897/2277-4343.07116>

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